A Case of Pleural Empyema Caused by *Streptococcus pneumoniae* Serotype 19A in a Fully Vaccinated Child

Tam Aşılı Bir Çocukta *Streptococcus pneumoniae* Serotip 19A’nın Neden Olduğu Bir Plevral Ampiyem Olgusu

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Abstract

*Streptococcus pneumoniae* is the most common cause of community-acquired pneumonia and empyema in children. After the childhood 13-valent pneumococcal conjugate vaccination programs, the incidence of invasive pneumococcal disease with vaccine serotypes has been decreased. However pleural empyemas caused by vaccine serotypes were reported in fully and appropriately vaccinated cases. Here in, we presented a case of pleural empyema and lung abscess caused by *S. pneumoniae* serotype 19A, in an immunocompetent and fully vaccinated child with 13-valent pneumococcal conjugate vaccine.

Keywords: Pleural empyema, *Streptococcus pneumoniae*, vaccine

Introduction

*Streptococcus pneumoniae* is the most common agent of community-acquired pneumonia and empyema in children. Isolating *S. pneumoniae* from normally sterile body fluids like the blood, cerebrospinal fluid and pleural fluid is defined as invasive pneumococcal disease (IPD). With the development of pneumococcal conjugated vaccine (PCV), the incidence of IPD with virulent serotypes in the vaccine has decreased (1). Primarily, 7-valent PCV (PCV7) containing serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F was started to be administered in the United States of America in 2000. In addition to 7-valent PCV, 10-valent PCV (PCV10) containing 1, 5, and 7F serotypes got license in Canada, Australia and Europe in 2008. Thirteen valent PCV included 3, 6A, and 19A serotypes in addition to PCV10, and obtained license from the European Drug Administration in 2009 (1). Thirteen valent pneumococcal conjugated vaccine (PCV13) was integrated to the expanded immunization program in Turkey in 2011, and vaccination rates reached 97% with PCV13 at the end of 2015 (1,2). Based on the fact that 13-valent PCV contains more serotypes, it possesses a broader protection and safety in proportion to PCV7 (3). Nevertheless, fully vaccinated children developing empyema with the serotypes in the vaccine have been reported. Here, a case of pleural empyema and
pulmonary abscess developed with *S. pneumoniae* serotype 19A in an immunocompetent, fully vaccinated child with PCV13 has presented.

**Case Report**

A previously healthy 3-year-8 months old female patient presented with complaints of malaise and cough lasting for 6 days and chest pain, respiratory distress, and tendency to sleep lasting for 3 days. Patient’s medical history, and her familial history was unremarkable. The patient was vaccinated with 4 doses of PCV13 according to the routine vaccination schedule. On her physical examination, body temperature was 37.1°C, respiratory rate was 100/min, hearth rate was 150/min, and oxygen saturation was 89%. Breath sounds of the left hemithorax were significantly decreased her expiration was prolonged, and rhonchus, suprasternal and intercostal retractions were present. Laboratory findings were as follows; hemoglobin: 11.3 g/dL, white blood cell count: 16.990/mm³ (82% neutrophil, 14% lymphocyte, 4% monocyte), thrombocyte count: 340,000/mm³, erythrocyte sedimentation rate (ESR) 99 mm/h, C-reactive protein (CRP): 270 mg/dL. Posteroanteior chest radiography demonstrated consolidation on the middle zone of the right lung, disseminated consolidation and pleural effusion extending to the apex on the left lung (Figure 1). The depth of the pleural effusion monitored on the left lung on thoracic ultrasoundography was measured as 10 mm. Diagnostic thoracentesis was performed and 90 mL purulent fluid was aspirated. Gram staining of the pleural fluid showed plenty of polymorphonuclear leucocytes, but no bacteria. The patient was started on empirically intravenous ceftriaxone (100 mg/kg/day, 2 doses), vancomycin (60 mg/kg/day, 4 doses), clindamycin (40 mg/kg/day, 4 doses) and oral oseltamivir (2 x 45 mg) since the patient admitted in the winter. The patient, who developed respiratory distress during follow up, was referred to the pediatric intensive care unit providing O₂ support with high flow nasal cannula (HFNC). On admission day 2, closed-tube thoracostomy drainage was undertaken and due to the fact that there were intense septations in the pleural space, tissue plasminogen activator (t-PA) was given into the pleural space. Tube thoracostomy catheter of the patient, who was given two doses of intrapleural t-PA and whose tube thoracostomy catheter was drained 470 mL of purulent fluid, was removed on admission day 9 due to the fluid drainage was terminated. While the blood culture of patient was negative, vancomycin-susceptible but penicillin, clindamycin and ceftriaxone-resistant *S. pneumoniae* was yielded on the pleural fluid culture of the patient. Clindamycin and oseltamivir treatment was terminated. The obtained isolate was typed/identified as serotype 19 by swelling of the capsule (Quellung reaction) with commercial antiserum belonging to the Statens Serum Institute (Copenhagen, Denmark). On thoracic ultrasonography performed after the removal of tube thoracostomy of the patient, a 70 x 37 mm in size solid formation, involving cystic-necrotic areas within, in the left lung was monitored. A thoracic computerized tomography (CT) was performed, and an appearance compatible with an abscess, approximately 80 x 55 x 55 mm in size, with peripheral contrast involvement and levelling within in the left lung lower lobe was detected (Figure 2). The patient was consulted by the department of thoracic surgery and monitoring with antibiotic treatment was decided...
Ceftriaxone treatment the patient was receiving was changed to meropenem on day 12 and vancomycin treatment was terminated on day 21. The abscess in the lower-mid zone of the left lung of the patient monitored with weekly thoracic ultrasonography was seen to have regressed to a size of 35 x 22 mm. Immunological evaluation demonstrated normal levels of total neutrophil and lymphocyte counts, serum IgG, IgA, IgM and IgE, lymphocyte subgroups and complement pathway. The patient did not have asplenia and HIV serology was negative. The patient, whose white blood cell count was 7280/mm³, ESR was 30 mm/h and CRP was 3 mg/dL in the tests performed on day 30 of meropenem treatment, was discharged with two doses of oral 7 mg/kg cefdinir treatment consecutively. In the tests done in the first month of polyclinic follow-up, patient’s white blood cell count was 7900/mm³, ESR was 11 mm/h and CRP was 3 mg/dL, and cefdinir treatment was stopped due to the fact that the abscess of the patient distinctly regressed. On thoracic CT of the patient, whose outpatient follow-up continued for three months, the abscess was found to have regressed completely. Written informed consent was received from the patient’s family in order for her medical information and imaging to be used in this study.

Discussion

A national, retrospective cohort study was conducted, in the United States of America after introduction of PCV7, to determine empyema incidence. Although empyema percentage related to pneumococcus decreased from 49% to 34%, a significant alteration did not occur in the incidence of pneumococcus-related empyema and hospitalization rate was found as 0.7/100,000 between the years 1997 and 2006. This increase in empyema despite the decrease in pneumococcus-related meningitis, bacteremia and pneumonia with 7-valent PCV has suggested that serotypes 1, 19A, 3 and 7, which are out of the coverage of this vaccine, are more frequently associated with empyema (5). In a study conducted in Canada comparing the incidence of invasive pneumococcal diseases in the pre-vaccination period, and post-PCV7 and post-PCV13 periods, it has been seen that post-vaccination hospitalization rates decreased from 17/100,000 to 4/100,000 and that age of admission with an invasive pneumococcal disease in the pre-vaccination period increased from 2 years of age to an average of 3.9 years after PCV13. It has been indicated that hospitalization after vaccination is much more related to focal infections (6). In a study carried out in our country, 156 children with empyema were analyzed in terms of S. pneumoniae serotypes irrespective of vaccination status, and S. pneumoniae has been detected as the agent in 53 (34%). S. pneumoniae serotypes were identified in 33 patients and serotypes 1 and 5 were found to be the most frequently observed ones (1). Serotype 3-associated pleural empyema cases have been seen after PCV13 vaccination in Greece and serotypes 19A and 3 have been reported as the most dominant serotypes, 2-3 years after the introduction of vaccine (7). In 2016, necrotizing fasciitis and acute purulent pericarditis complicated with bronchopleural fistula, pneumothorax and subcutaneous empyema caused by S. pneumoniae serotype 19A in a 4-year-old fully vaccinated child was reported, and again in 2017, a case that developed pleural empyema due to serotype 19 was reported in a 2-year-old child from Casablanca (8,9). In a study of 161 pediatric cases in the United Kingdom, vaccine failure was found to be 0.19/100,000 in those vaccinated with PCV7; and 0.66/100,000 in those vaccinated with PCV13 and it was reported that vaccine failure was rare, however bacteremic lower respiratory tract infection and empyema were more frequent with serotypes added to KPA13 compared to KPA7 serotypes (10). Vaccination failure is a state where the individual cannot produce antibodies against the vaccine after primary or booster doses. The causes of vaccination failure can be related to the patient, the vaccine or vaccination. Patient related causes can be arisen from, due to immunodeficiency, being unable to produce antibody against vaccine itself, components or serotypes of the vaccine. Moreover, patient age, the presence or absence of nutrition deficiency, whether or not the patient was healthy at the time of vaccination, and whether or not the exposure to the pathogen before being vaccinated, effect the individual’s response to the vaccine. Causes related to the vaccine itself are; first the vaccine does not include some serotypes and genotypes, second antigenic interaction and production stage related reasons. Vaccination-related causes are conditions such as mistakes made during vaccine administration, not conforming to cold chain during storage and transportation, and failure in following the vaccination schedule (4). Our case was fully vaccinated with PCV13 and 32 months had passed from the booster dose. Immunodeficiency examinations of our case, for whom a sufficient amount of time had passed for the protective immunological response to develop, was normal and IgG titer against pneumococcus was at a protective level. Up-to-date treatment approach to pleural empyema and pulmonary abscess is tube thoracostomy and underwater seal drainage in addition to antibiotic therapy. Recovery frequency of the cases with pulmonary abscess with medical treatment without a sequel has been reported as 70%-90%, and thus, open surgery that has a high rate of mortality is preferred in a very limited number of cases (11). Our case was successfully treated with tube thoracostomy and underwater seal drainage together with antibiotic treatment.

In conclusion, parapneumonic pleural empyema may develop with the serotypes within the vaccine in immunocompetent individuals fully vaccinated with PCV13. Therefore, even though national vaccination schedule includes PCV13,
monitoring the surveillance of invasive pneumococcal diseases and defining agent serotypes have importance.

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**References**


